# Acute Kidney Injury Secondary to Lonomia envenomation. Case Report and Literature Review

Lesión renal aguda secundaria al envenenamiento de Lonomia. Informe de casos y revisión de literatura

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Daniel Martín Arsanios Internal Medicine Resident, La Sabana University, Chía, Colombia Elías Josué Quintero Muñoz MD, La Sabana University, Chía, Colombia Nicolás David Santoyo Sarmiento<sup>a</sup> MD, El Bosque University, Bogotá, Colombia Álvaro Arteaga MD, La Sabana University, Chía, Colombia María José Olaya MD, La Sabana University, Chía, Colombia Aleiandra Perilla Rodríguez MD, Universidad Cooperativa de Colombia, Villavicencio, Colombia Alirio Batidas Goyes MD. Internist, nuemologist, epidemiologist, La Sabana University, Chía, Colombia

<sup>a</sup> Autor de correspondencia: santoyosarmienton@gmail.com

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#### ABSTRACT

Introduction: The envenomation by Lonomia species mainly occur in rural areas. The main symptom of this condition is an alteration in the clotting times and hemorrhage. However, acute kidney injury is been also identified in some case reports. Therefore, it should not be disregarded and to understand the pathophysiological mechanism, diagnosis and treatment of this clinical feature results necessary in order to expand awareness and to prevent further medical complications and death. Materials and Methods: We present a case report and scoping review of the literature to characterize patients with acute kidney injury secondary to Lonomia envenomation by a case report and describing the pathophysiology, diagnosis, treatment and prognosis of patients suffering from this condition. The information was obtained from 4 databases: Medline, Embase, LILACs and Ovid. We used the following search strategy: ((("Acute Kidney Injury" [Mesh]) AND "lonofibrase" [Supplementary Concept]) OR lonomy accident) OR lonomia. Results: We identified ten case reports of Lonomia spp. Envenomation, nine of them were caused by L. obliqua and one by L. achelous. A cohort study that included 37 patients who were exposed to the L. obliqua venom was also included. All patients were adults, lived in the countryside and developed acute kidney injury. Conclusions: Acute kidney injury has been identified as one important clinical feature

of the *Lonomia* envenomation. The development of this condition is mediated by various pathophysiological mechanisms. To perform an adequate diagnosis, coagulation times, fibrinogen levels, hemogram, kidney function and urinalysis should be done. The definitive treatment depends on the type of envenomation. The prognosis for these patients is not well described yet. However, our patient had a good response to the described management given.

#### Keywords

Lonomia obliqua ; caterpillar; acute kidney injury; poisoning.

#### RESUMEN

Introducción: El envenenamiento por Lonomia se produce principalmente en las zonas rurales. El síntoma principal de esta afección es hemorragia y una alteración en los tiempos de coagulación. Sin embargo, la lesión renal aguda también se ha identificado en algunos informes de casos. Por lo tanto, no debe ser ignorado y para entender el mecanismo fisiopatológico, diagnóstico y tratamiento de esta característica clínica resulta necesario ampliar el conocimiento y prevenir las complicaciones médicas. Materiales y métodos: Presentamos un informe de caso y una revisión de la literatura para caracterizar los pacientes con lesión renal aguda secundaria al envenenamiento por Lonomia por medio de la descripción de la fisiopatología, diagnóstico, tratamiento y pronóstico de pacientes que sufren de esta afección. La información se obtuvo de 4 bases de datos: Medline, Embase, LILACs y Ovid. Usamos la siguiente estrategia de búsqueda: ((("Lesión renal aguda" [Malla]) Y "lonofibrase" [Concepto suplementario]) O accidente lonómico) O accidente por lonomia. Resultados: Identificamos diez informes de casos de envenenamiento por Lonomia spp., nueve de ellos fueron causados por L. obliqua y uno por L. achelous. También se incluyó un estudio de cohorte que incluyó a 37 pacientes que estuvieron expuestos al veneno de L. obliqua. Todos los pacientes eran adultos, vivían en el campo y desarrollaban lesión renal aguda. Conclusiones: La lesión renal aguda se ha identificado como una característica clínica importante del envenenamiento por Lonomia. El desarrollo de esta está mediado por varios mecanismos fisiopatológicos. Para realizar un diagnóstico adecuado, se deben realizar tiempos de coagulación, niveles de fibrinógeno, hemograma, función renal y análisis de orina. El tratamiento definitivo depende del tipo de envenenamiento. El pronóstico para estos pacientes aún no está bien descrito. Sin embargo, nuestro paciente tuvo una adecuada respuesta.

#### Palabras clave

Lonomia obliqua; Caterpillar; Lesión renal aguda; Envenenamiento.

#### Introduction

Envenomation by venomous animals is a public health problem, and the morbidity and mortality associated with cases of envenomation by different animals has an impact on the population and the Colombian health system.

The first records of accidents due to contact with caterpillar spines are found in Brazil in 1912 and were described by Zoroastro. These accidents occur mainly in rural areas, and most often affect the upper extremities of patients (1,2). The increase in accidental envenomation with caterpillar venom is associated with deforestation and the secondary elimination of natural predators (2,3).

The main symptom of *Lonomia* envenomation is an alteration in the clotting times associated with a consumption coagulopathy that leads to hemorrhagic symptoms (1-4). Additionally, it has been described that the venom causes nephrotoxicity and a broad clinical spectrum of deterioration of renal function in both human beings and in vitro studies (3). Although secondary coagulopathy is what occurs most frequently, acute kidney injury is one of the leading causes of death by envenomation. In 1980, 18% of patients developed acute kidney injury, and 50% of them died, but there was an incidence of less than 5.2% (3).

Despite the low incidence of cases of acute kidney failure, it is necessary to understand the pathophysiological mechanism, diagnosis and treatment of this complication to expand awareness and reduce acute kidney failure as a progression of envenomation, and death.

#### Case Report

An 80-year-old male patient from Puerto Gaitán (Meta-Colombia), with no relevant medical history, who presents a clinical course of 4 days of evolution, consisting of pain in the left flank and macroscopic hematuria subsequent to contact with poisonous caterpillar (Figure 1). On admission, the patient is hemodynamically stable, with pain on palpation of the left flank, without signs of peritoneal irritation without any other positive finding. The evolution of the laboratories is seen in Table 1. After clinical and paraclinical following and improvement of the patients, discharge occurs.

#### Figure 1.

Poisonous caterpillar of Lonomia spp. with their characteristic bristles



### Table 1.

Follow-up of the patient's laboratories results since admission

Laboratories	Admission	2 days after admission	Post antilonomic serum administration	One day after antilonomic serum	
Creatinine	1.08 mg/dl	1.24 mg/dl	0.99 mg/dl	1.15 mg/dl	
Blood urea nitrogen (BUN)	22.7 mg/dl	27.1 mg/dl	21.2 mg/dl	23.9 mg/dl	
Prothrombine time (PT)	3 min	19.3 s	17.4 s	14.4 s	
Partial thromboplastin time (PTT)	37.3 s	35.8 s	28.9 s	20.8 s	
Fibrinogen	92.5 mg/dl	92.5 mg/dl	77 mg/dl	157 mg/dl	

### Methodology

We carried out a scoping review in the Pubmed, Embase, OVID, and LILACS databases. The search strategy was performed using the terms: "Acute Kidney Injury" [Mesh]) AND "lonofibrase" [Supplementary Concept]) OR lonomia accident) OR lonomia. Articles published from January 1, 1998 to November 25, 2018 were include. We included case reports, case series, cohort studies, cases and controls, experimental and cross-sectional studies in English, Spanish, Portuguese, French and Italian that included patients older than 18 years with acute kidney injury, defined as an increase in serum creatinine  $\geq$  0.3 mg/dL in 48 hours or an increase in serum creatinine  $\geq 1.5$  times the baseline value or a urine output < 0.5 ml/kg/h for 6 hours (5) secondary to *Lonomia* envenomation. We excluded articles in which creatinine or urine output values were not documented. The search was conducted separately by two researchers, DMA and MJO, using the search strategy described above. 1 describes the articles included and excluded by title, abstract and finally in the complete revision of the same.

### Results

We identified ten patients reported to have be envenomed by L. spp. and one cohort study that included 37 patients who were exposed to the L. obliqua venom. All of them were ranged between 26 and 70 years of age. All the cases were reported to have happened in rural areas. In nine patients, L. obliqua was identified as the cause of envenomation, and only in one case it was due to L. achelous. The common clinical features of these patients at the arrival of the emergency department were the acute kidney injury and the alteration in the clotting times with secondary hemorrhagic manifestations of different localizations. Only one patient was pregnant at the time of exposure to the venom who performed with placental abruption. The summary of the included studies is seen in Table 2.

### Table 2.

Summary	of the	eleven	included	studies
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Author/Date	Type of study	Patients	Sex	Age	Type of Lonomia	Clinical picture	Complications	Treatment
Sano-Martins et al. (2017)	Case series	14	NE	NE	Lonomia obligua	Increase in creatinine up to 2.7 mg/dL, normalization at 3 days	NE	NE
Corrên et al. (2004)	Case report	1	м	44	Lonomia app	Increase in creatinine up to 18.5 mg/dL, normalization at 41 days	Severe hemorfhagic syndrome	Transfusion 600 nd RBC, 10 Unit platelets Dialysis
Hui Wen et al. (1998)	Case report	1	м	37	Lononia obligua	Increase in creatinine up to 10.1 mg/dL, 36 days after creatinine of 3.8 mg/dL	Severe hernorthagic syndrome	Hemodialysis
Sanchez et al. (2015)	Case sories	1/6	м	46	Lonomix obligua	Increase in creatinine up to 1,5 mg/dL in 72 hours	Hemorrhagie syndrome, Mild hepotic failure	Anti-Ionomic serum, Coeticoid, Vitamin k
Riella et al. (2008)	Case report	1	,	68	Lonomáx obligua	Increase in creatinine up to 8.2 mg/dL, then 4.45 mg/dL and 1.75 mg/dL at 9 days and 5 meeths	Hemorihagic syndrome	Anti-Ionomic serum Hemodialysis Transfusion 1 Unit RBC, 3 Unit Cryoprecipitate
Kowaca (2006)	Case report	1	F	70	Lonomia obligua	Increase in creatining from 0.8 mg/dL to 1.1 mg/dL, the patient died	Sudden coma Hemorihagic syndrome	NE
Malaque (2006)	Case report	1	м	64	Lonomia obligua	Increase in creatinine up to 6.4 mg/dL, then 5.6 mg/dL and 1.5 mg/dL at 20 and 30 days	Hemorihagic syndrome	Anti-Ionomic scrum Dialysis
Camborgi (2006)	Cahort	39	MŦ	-	Lonomia obligua	Croatinine >1.5 mg/dL in sill cases. 16 required hermodialysis 4 did not regain renal function post- dialysis and ALS	Hemorihagic syndrome'	Anti-Ionomic servan Diadysis
Cardenas (2002)	Case report	1	,	52	Lonomia obligua	Baseline creatinine 12.1 mg/dL, 20 days after creatinine of 2.9 mg/dL	Hemserhagie syndrome	Presh forzen plasma, Cryoprecipitate Hemodialysis
Pineda (2001)	Case series	1	F	26	Lonomia Achelous	Baseline creatinine 9 mg/dL, the patient died	Hemorrhagic syndrome	Transfusion RBC, platelets Continuous hemodiafiltration
Schmitherger P (2013)	Case report	1	м	60	Lonomia obligua	Edematous syndrome, arthralgias and amaria. Baseline creatinine of 7.3 mg/dL, day 16 creatinine 11.9 mg/dL	End-stage renal disease	Anti-Ionomic scram and hemodialysis

DANIEL MARTÍN ARSANIOS, ELÍAS JOSUÉ QUINTERO MUÑOZ, NICOLÁS DAVID SANTOYO SARMIENTO, ET AL.

#### Discussion

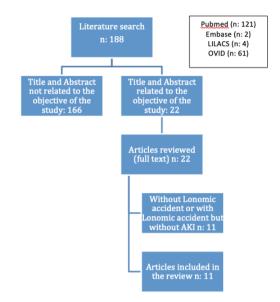
The patient presented acute kidney injury (AKI) I according to laboratory results associated with countless coagulation times with decreased fibrinogen without platelet decrease, a picture compatible with *L. obliqua* poisoning according to the pathophysiological study of Alvarez Florez et al., in 2010. (6).

We found 1 cohort study and 10 case reports in which patients with lonomiasis developed acute kidney injury. In all cases there was alteration in the clotting times at the time of admission to the hospital. The patients presented hemorrhagic syndrome and AKI between 8 and 72 hours after contact with the venom (7), also hematuria and mucocutaneous bleeding. AKI was determined using the current creatinine elevation criteria (which were common to all cases). It should be noted that among the cases not selected for not presenting AKI, other potentially fatal bleeds were found, 9% of which were gastrointestinal, 4% intracranial, and 4% alveolar hemorrhage.

Among the cases in which AKI was documented, 9 were caused by *L. obliqua*, 1 by *L. achelous*, and one was not typified. These figures are consistent with the epidemiological data in the literature, which shows a higher incidence of serious clinical involvement due to *L. obliqua* venom in Colombia. Figures 2 and 3 (7).

#### Figure 2.

Flow diagram of the criteria to consider in the researching process



#### Figure 3.

Physiopathology of the acute renal injury secondary to Lonomia envenomiaton



Regarding the pathophysiology of the kidney injury, in most patients it was related to the hemodynamic changes secondary to the hemorrhagic syndrome. Among the most important changes are the decrease in systemic blood pressure and prerenal injury secondary to generalized renal ischemia due to a state of hypoperfusion (8). In addition, kidney local inflammatory processes caused by the larva venom toxins attribute an essential complementary factor to the renal involvement (3). A renal puncture with biopsy was performed in one of the reported cases and on one of the patients included in the cohort study. It showed acute tubular necrosis with mesangial proliferative glomerulonephritis and mononuclear interstitial infiltrates (9), and only signs of acute tubular necrosis (3), respectively, which supports the previous approach.

The case report and the theme review by Ávila et al., in 2013, and the Brazilian Ministry of Health agree that the patient involvement can be classified as mild, moderate or severe (7,8). According to this, all the cases reported and those included in the cohort study had a severe involvement due to the presence of AKI.

According to the creatinine elevation for the diagnosis of acute kidney injury, the recorded range ranged between 1.1 and 12.1 mg/dL. Regarding the AKIN classification for AKI, one of the patients was AKIN I, two cases corresponded to AKIN II, and seven were AKIN III. Within the reported cases, seven patients were taken to hemodialysis and one developed chronic kidney failure. Among the clinical and paraclinical characteristics for renal replacement therapy, it was observed that all patients had a creatinine level  $\geq 4 \text{ mg/dL}$  and clinical signs of uremia; 71% had persistent oligo-anuria for more than 48 hours, and 14% had hyperkalemia (potassium of 5.7 mEq/L). Of the 11 patients, 4 received Anti-Lonomic Serum (ALS). The duration of dialysis therapy in these patients did not exceed 20 days, and there was clinical and paraclinical improvement in the control performed the day after the administration of the serum, especially in the hematological parameters and the coagulation.

On the other hand, of the 2,067 patients included in the cohort study, 39 presented AKI, defined in this study as: serum creatinine greater than 1.5 mg/dL without a history of kidney disease. However, it is important to mention that the study was carried out in two different periods, because before 1995 there was no specific treatment for the lonomic accident; due to this, the initial management of the first 175 patients who entered the cohort was different. Of these, 5 developed AKI (2.9%). On the other hand, of the 1,892 patients who later entered the study, and who were treated with ALS, 34 had AKI. In both periods there were two patients who progressed to chronic kidney disease (10.3%), and there was only one fatal outcome due to AKI in the first period (3).

It is worth noting that this study suggests that the development of AKI is directly related to the amount of inoculum with which the patients had contact, and this, in turn, is proportionally linked to the number of caterpillars at the time of exposure. 90% of the patients with AKI recovered all or part of the renal function, and these cases were caused by *L. obliqua* (3).

Considering the case reports and the cohort study, the duration of dialysis therapy generally varied according to the decrease in serum creatinine values over time and the progressive increase of urine output; four days was the shortest time interval, and 45 days the longest.

In many cases, symptomatic treatment was given with corticosteroids and antihistamines, given the inflammatory and anaphylactic manifestations presented by the patients. However, the treatment for erucism consists of washing the affected area with cold water, applying cold compresses, infiltration with 2% lidocaine and topical corticosteroids (10).

Among the treatment alternatives for the moderate to severe involvement found in the literature and that were thought effective before ALS became available, are antifibrinolytic drugs (aprotinin, epsilon-aminocaproic acid and tranexamic acid), purified fibrinogen, cryoprecipitates, and replacement therapy (whole blood or fresh frozen plasma) (7,8). However, only a small benefit has been seen in the resolution of symptoms, but not a total reversal of the effects of the larva venom toxins (7).

With the production and implementation of ALS, not only a rapid and evident resolution of the clinical manifestations was observed, but a clear reversal of the effect of the venom toxins due to its main component, which is a heterologous immunoglobulin specific to the *L. obliqua* venom. (11), as long as the treatment was started within 48 hours after exposure to the venom (12). As previously mentioned, not all patients received ALS. However, there were no fatalities among those who received

it. Instead there was a higher mortality in those who received alternative treatments, which increased when patients were exposed to the *Obliqua* species. The cause of death in these cases was secondary to the manifestations of the hemorrhagic syndrome, especially to intracranial hemorrhages (2).

Finally, regarding the prognosis of patients with lomonic accident associated with AKI, it has been seen that the degree of morbidity and mortality is related to the severity of the clinical involvement and the alterations in the paraclinical tests at the time of hospitalization (12). Regarding the associated kidney condition, the follow-up of some of these patients was done with periodic controls of serum creatinine levels to define the condition's progression or improvement, given that some patients developed chronic kidney disease after the lomonic accident. Some of the risk factors identified in the cohort study were age, time elapsed before starting treatment, occupation, number of caterpillars involved, severity of bleeding and low platelet count. However, there are no conclusive studies on how the outpatient control of these patients at risk of developing chronic kidney disease should look like (3).

## Pathophysiology

Envenomation by Lonomia spp., especially by L. obliqua, causes acute kidney failure through various mechanisms like massive deposition of microthrombi in glomeruli associated with disseminated intravascular coagulation, direct nephrotoxic action of the venom, hemodynamic changes, ischemic renal lesions, band acute tubular necrosis following the hemorrhagic shock (6), that will be describe below. Since the kidney is a highly vascularized organ whose function is to filter and excrete waste, including venom, it is very susceptible to toxins, with a variable spectrum of manifestations ranging from interstitial tubule nephritis, passing through glomerulonephritis, vasculitis, tubulointerstitial nephritis and acute tubular necrosis, the latter being the most frequent (13). Envenomation

with this species causes what is known as severe hemorrhagic syndrome, which has clinical manifestations such as coagulation alterations, ecchymosis, acute kidney failure and generalized hemorrhage (2,9,13,14). The components with pro-coagulant, fibrinogenolytic, proteolytic and hemolytic activity cause a condition compatible with disseminated intravascular coagulation associated with a consumption coagulopathy, caused by a plasminogen activator, an enzyme similar to plasmin, two proteases that act on factor V and another on factor XIII, generating an activation in the fibrinolytic cascade that can mask the latter (4,15), which are one of the mechanisms that cause kidney damage (14,16,17) by forming microthrombi in the kidney microcirculation and generating a state of hypoxia and tissue ischemia that alters its functioning and causes cell death (6,3,14). On the other hand, the resulting hemolysis generates hemoglobin deposits in the renal tubules, and its degradation produces oxidative stress and endothelial damage (18).

However, the main mechanisms described for progression to acute kidney failure are the hemodynamic changes caused by the hemorrhagic syndrome, which decrease the mean arterial pressure, and therefore the renal perfusion pressure, which leads to a decrease in the glomerular filtration rate; hemolysis, which generates waste products that alter kidney function; alteration of vascular permeability, which causes edema, tubular obstruction and glomerular fibrin deposition, which contributes even more to tissue damage and becomes a postrenal etiology of this acute kidney failure; and last but not least, the direct nephrotoxicity of the venom produced by the action of metalloproteinases, serine proteinases, C-type lectin, phospholipases A2 and different oxidases (12, 19, 20).

A decrease in the glomerular filtration rate is observed, associated with the inability of renal tubules to absorb electrolytes due to the damage caused in the tubular epithelium; this alters the function of the Sodium-Potassium ATPase pump (21), initially increasing its function, which initially increases urine output and decreases urine osmolarity. After 6 hours of exposure to the venom, massive proteinuria is observed. Histologically, there is acute tubular necrosis with tubule dilatation, cytoplasmic vacuolization, degeneration and desquamation of the necrotic epithelium, which causes an intense inflammatory response with tissue fibrosis at 48-96 hours (12,14,17).

The kinin-kalicrein system (KKS) is activated by the kalicrein- and kininogenase-activating proteins present in the venom, which generates the release of bradikinins (pro-inflammatory substance) by means of low molecular weight kininogen (LMWK) and stimulating precalikrein in plasma (14); this leads to an increase in vascular and tissue permeability resulting in a decrease in blood pressure and edema in peripheral tissues (14,16,22). The KKS is important for the pathophysiological mechanism of renal failure, the kalikreina directly activates factor XII which generates an activation of Kinina for thrombus formation, by the intrinsic pathway (14,15). The effect of quinines is found in the bradykinin 1 (BR1) and bradykinin 2 (BR2) receptors, these receptors are involved in different processes such as inflammation, atherosclerosis, neuropathic pain and cerebrovascular disease; but in experimental models, when there is a blockage of the KKS, a decrease in flow, capillary leakage, inflammation and thrombi has been evidenced decreasing the deleterious effects of poisoning and with it the renal function (14). The venom components can be located in vascular smooth muscle cells (VSMC) in vivo and in vitro, this activation contribute to induce the formation and deposition of fibrin in the glomerular capillaries, and then reduce the filtration capacities, the L. obliqua venom can trigger reactive oxygen species (ROS) through the activation of NADPH oxidase favoring the coagulation cascade and incoagulability (14).

It has been seen that kidney tissue undergoes an inflammatory process caused by *Lonomia obliqua* venom, associated with a systemic inflammatory response whose main symptoms are pain and edema at the contact site (23,24) with a systemic production of TNF, IL8, IL6, histamine, prostaglandins, nitric oxide (NO) and other vasoactive inflammatory mediators or endothelial adhesion mediators. At kidney level there is also a series of inflammatory reactions; there is an increase in leukocyte migration and cell adhesion, production of IL8, TNFa, ICAM-1, E-selectin and nitric oxide, and activation of COX-2, iNOS, NADPH oxidase (6 ,9,14,16,25,26); this produces oxygen reactant of mitochondrial origin, which further damages endothelial cells, thus inducing the adhesion and extravasation of all this inflammatory reaction, increased vascular permeability, glomerular dilatation, distention of Bowman's space and interstitial edema. In addition, inhibition of metalloproteinases (MMP) has been observed, which produces deposits of collagen and other matrix proteins (14,24,27). The Figure 3 describe the physiopathology.

### Diagnosis

The clinical record and physical examination are essential factors for an adequate diagnosis. Patients usually consult after a not very long period of evolution after the contact with the larva of L. obliqua (28,29), reporting a clinical picture of malaise associated with burning pain at the contact site, myalgia, and occasionally high temperature, which is usually accompanied hemorrhagic systemic manifestations bv (gingivorrhagia, epistaxis, ecchymosis and even digestive hemorrhages) (8,15,17,30). In some very severe cases there may even be intracerebral hemorrhage. However, the prognosis is guarded, and this is the first cause of death in these patients (16). To make an adequate diagnosis, paraclinical tests must be performed; an essential test are the coagulation times, which are altered in all cases with an important prolongation, secondary to a hemorrhagic process with a fibrinolytic pathophysiological basis. Prothrombin time (PT) and partial thromboplastin time (PTT) values are usually variable, due to the initial characteristics of the patient's coagulation system (10,11,31). However, it has been observed that the values

in *L. obliqua* envenomation are always above the control, since the larva has two procoagulant proteins that affect the coagulation cascade in the common pathway, lopap prothrombin activator, and losac factor X activator; this is the reason why both PT and PTT are altered (13,32,33). It is possible to evaluate at the same time factors that are altered by wear, such as fibrinogen and platelets, and there is significant hypofibrinogenemia and thrombocytopenia secondary to the intrinsic fibrinolytic hemorrhagic process that develops with L. obliqua toxins (16,34). Normal volume anemia may also occur due to blood loss, which will be directly proportional to the patient's clinical condition (34,35).

A kidney function test and a partial urinalysis should be performed, as in this case hematuria is a clinical manifestation of an intrinsic kidney disorder that is associated with the development of acute tubular necrosis in some patients, which is the basis of acute kidney failure, and therefore of the development of an unsatisfactory systemic state with high mortality rates; this factor alerts to the need of establishing a fast and effective therapeutic plan (9,28,36,37). The diagnosis of acute kidney failure was made in a study by Gambordi et al. (3), with creatinine results >1.5 md/dL in patients without a history of kidney failure (3); the study reported that, although the administration of ALS did not produce an immediate improvement in acute kidney failure, it did have an impact on the progression of the disease in terms of its severity and the prevention of chronic kidney disease (3). Another diagnostic method performed by Burdmann et al. (1996) was a renal biopsy 17 days after the caterpillar envenomation; the histopathology showed thickening of Bowman's capsule and tubule atrophy; other findings in the renal biopsy were compatible with acute tubular necrosis in the recovery phase (3). However, coagulopathy makes it difficult to perform this study in the acute phase. Proteinuria was also observed in patients with acute kidney failure, which is greater in the first 6-12 hours after the contact (3).

### Treatment

Initially, a symptomatic therapeutic approach can be effective in terms of reducing pain, normalizing temperature curves and reducing bleeding, with all the hemodynamic implications that this entails. For this reason, the use of NSAIDs, corticosteroids and first-generation antihistamines is indicated to decrease the inflammatory response and achieve an initial symptomatic stability (37).

As previously stated, the pathophysiology of *L. obliqua* envenomation consists of the development of consumption coagulopathy due to the inoculation of two procoagulant enzymes present in the larva venom, which generate a decompensated response by the fibrinolytic system. Therefore, antifibrinolytic agents do not produce a symptomatic or laboratory improvement; they inhibit the degradation of fibrin and do not produce an improvement in cases of coagulopathy in which the factors are consumed, as in the *L. obliqua* envenomation (15,38).

Conversely, in cases of *L. achelous* envenomation, the symptomatology is secondary to an abnormal and exaggerated fibrinolytic activation (39), so the use of antifibrinolytic drugs such as aminocaproic acid and tranexamic acid is recommended, in order to decrease the bleeding rate, which in some cases can be severe; this improves the patient's hemodynamic conditions, preserving his stability and reducing the risk of developing acute kidney failure due to renal hypoperfusion with a subsequent decrease in the glomerular filtration rate (31,40). This implies a clinical and symptomatic improvement in cases of *L. achelous* envenomation.

However, in cases of *L. obliqua* envenomation, the larva toxins continue to cause systemic damage if the antidote or anti-lonomic serum, also known as "anti-venom", is not administered (41); this antidote was developed for the first time in 1996 in Brazil, by extracting fragments of immunoglobulin G from previously immunized horses (38). The administration of this serum produces a significant reversal in the clinical condition of the patient envenomed by *L. obliqua*; a symptomatic and paraclinical improvement is observed, since it has been shown that a few hours after administering this serum, the clotting times are normalized, there is a substantial increase of the fibrinogen and in the platelet count, and optimal values are observed in kidney function tests; this is secondary to the increase in renal perfusion, with a subsequent increase in the glomerular filtration rate and a decrease in the nephrotoxicity produced by toxins inoculated by the larva-man contact (6,15,38).

In the same way, the use of antibiotics, specifically of first-generation cephalosporins, has been described to prevent or treat the concomitant development of skin and soft tissue infection (42). It is also recommended to give vitamin K to these patients, as this will enhance the action of vitamin K-dependent coagulation factors, which promotes an adequate homeostatic control (42).

Recently, in an experimental study in animals conducted by Berger et al. (14), they describe the inhibition of KKS by means of aprotinin as a complementary management to ALS or as an alternative to reduce the complications and effects of poisoning through the inhibition of receptor BR1 and BR2 (14).

## Prognosis

Although epidemiologically significant morbidity and mortality rates have not been described in cases of *L. obliqua* envenomation, it has been found that the prognosis of the patients is directly related to the symptoms and the initial clinical and laboratory conditions with which the patient consults for the first time. The presence of intracerebral hemorrhage as a clinical manifestation of consumption coagulopathy has fatal outcomes, which is the first cause of death in these patients (16,36).

Similarly, patients with acute kidney failure as a manifestation of nephrotoxicity from the larva have hemodynamic and systemic repercussions that can quickly lead to multiorgan dysfunction (9,36). Intravascular hemolysis is a rare complication that has been observed only in the most severe cases of this pathology, associated with contact with a large number of caterpillars, and as well as the two aforementioned entities, it also has a worse prognosis in those patients who present them (43,44).

The patient's degree of hemorrhage, and even more so his degree of shock, warns of the need for an adequate approach and optimal hydric resuscitation, as well as for the rapid administration of pro-hemostatic drugs, such as antifibrinolytic drugs and ALS (40).

## Conclusions

Even though the most frequent clinical feature of the *Lonomia* envenomation is the alteration of clotting times, it has been found that acute kidney injury should not be disregarded as this condition increases the risk of further undesirable outcomes or even death.

The development of acute kidney injury is mediated by various mechanisms. These include the hypoxia and tissue ischemia caused by the haemodynamic changes and the microthrombi formation secondary to the haemorrhagic syndrome. Moreover, a proinflammatory state appears to affect the kidney locally, not only worsening its perfusion but also decreasing progressively the glomerular filtration rate and making it prone to develop acute tubular necrosis.

To perform an adequate diagnosis, some of the tests that should be done are the coagulation times, fibrinogen levels, an hemogram (searching for thrombocytopenia and anemia) in junction with the kidney function and urinalysis. Although some findings in the renal biopsy are helpful to confirm the diagnosis, this procedure is not feasible in the acute phase of the disease and could delay treatment.

A symptomatic approach for treatment is indicated to decrease pain and the inflammatory response. However, the definitive manage depends on the type of envenomation. For *L. obliqua* species, the anti-lomonic serum is the first line treatment to control promptly the haemorrhagic manifestations while for *L*. achelous, the antifibrinolytic agents may perform well.

Finally, the prognosis for these patients is not well described yet. Nonetheless, it is proportional to the severity of the clinical findings, the localization of the bleeding and the time of setting the treatment. In the case report, our patient had a good response to the treatment given as seen in his clinical and laboratory progression.

## References

1. Medeiros DNM, Torres HCC, Troster EJ. Accident involving a 2-yearold child and *Lonomia obliqua* venom: clinical and coagulation abnormalities. Rev Bras Hematol Hemoter [Internet]. 2014;36(6):445–7. Available from: htt p://www.sciencedirect.com/science/art icle/pii/S1516848414001169

2. Riella MC, Chula D, de Freitas S, Mazza MM, Pachaly MA. Acute renal failure and haemorrhagic syndrome secondary to toxin of caterpillars (*Lonomia obliqua*). Clin Kidney J [Internet]. 2008 Dec 1 [cited 2018 Nov 25];1(6):445–6. Available from: https://academic.oup.com/ckj/ar ticle-lookup/doi/10.1093/ndtplus/sfn1 38

3. Gamborgi GP, Metcalf EB, Barros EJ. Acute renal failure provoked by toxin from caterpillars of the species *Lonomia obliqua*. Toxicon [Internet]. 2006 Jan [cited 2018 Sep 21];47(1):68–74. Available from: http://www.ncbi.nlm.n ih.gov/pubmed/16310819

4. Arocha-Piñango CL, Guerrero B. Lonomia obliqua and haemorrhagic syndrome. Lancet. 1999; 354:1304. Available from: https://www.ncbi.nlm. nih.gov/pubmed/1052066646

5. Khwaja A. KDIGO Clinical Practice Guidelines for Acute Kidney Injury. Nephron. 2012 Aug;120(4):c179–84.

Alvarez Flores MP, Zannin 6. M. Chudzinski-Tavassi AM. New insight into the mechanism of Lonomia obliqua envenoming: toxin involvement and molecular approach. Pathophysiol Haemost Thromb [Internet]. 2010 [cited 2018 Sep 30];37(1):1–16. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/2071412642

7. Ávila A, Moreno A, Garzón A, Gómez A. Accidente lonómico Lonomiasis. Acta Médica Colomb. 2013;38:95–100.

8. Sarmiento NS, Arsanios DM, Quintero Muñoz E, Santoyo Sarmiento C. Lonomic accident and allergic reaction to anti-lonomic serum: case report and literature review [Internet]. 2018 [cited 2018 Sep 13]. Available from: http://www.ijcrls.com

9. Berger M, Santi L, Beys-da-Silva WO, Oliveira FMS, Caliari MV, Yates JR, et al. Mechanisms of acute kidney injury induced by experimental *Lonomia obliqua* envenomation. Arch Toxicol [Internet]. 2015 Mar 6 [cited 2018 Sep 30];89(3):459–83. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/24798088

10. Schmitberger P, Fernandes T, Santos R, de Assis R, Gomes A, Siqueira P, et al. Probable chronic renal failure caused by Lonomia caterpillar envenomation. J Venom Anim Toxins Incl Trop Dis [Internet]. 2013 Jun 3 [cited 2019 Apr 7];19(1):14. Available from: http://jvat.biomedcentral.com/ar ticles/10.1186/1678-9199-19-14

11. Da Silva WD, Rocha Campos ACM, Gonçalves LRC, Sousa-E-Silva MCC, G. Higashi H, K. Yamagushi I, et al. Development of an antivenom against toxins of *Lonomia obliqua* caterpillars. Toxicon [Internet]. 1996;34(9):1045–9. Available from: ht

### tp://www.sciencedirect.com/science/ar ticle/pii/0041010196000529

12. Sano-Martins IS, Duarte AC, Guerrero B, Moraes RHP, Barros EJG, Arocha-Piñango CL. Hemostatic disorders induced by skin contact with *Lonomia obliqua* (Lepidoptera, Saturniidae) caterpillars. Rev Inst Med Trop Sao Paulo [Internet]. 2017 Apr 20 [cited 2018 Mar 19];59:e24. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/28443942

13. Chudzinski-Tavassi AM, Schattner M, Fritzen M, Pozner RG, Reis CV, Lourenço D, et al. Effects of lopap on human endothelial cells and platelets. Pathophysiol Haemost Thromb [Internet]. 2001 [cited 2018 Sep 30];31(3–6):257–65. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/11910193

14.Berger M et al. Renal and vascular effects of kallikrein inhibition in a model of Lonomia obliqua venominduced acute kidney injury. PLOS Negl Trop Dis. 2019;13(2). https:// doi.org/10.1371/journal.pntd.0007197

15.Sano-Martins I, Gonzalez C. Effectiveness of Lonomia antivenom in recovery from the coagulopathy induced by Lonomia orientoandensis and Lonomia casanarensis caterpillars in rats. PLOS Neglected Tropical Diseases. August 16, 2018. Available from: https://doi.org/10.1371/journal.p ntd.0006721

16.Pinto AFM, Berger M, Reck J, Terra RMS, Guimarães JA. Lonomia obliqua venom: In vivo effects and molecular aspects associated with the hemorrhagic syndrome. Toxicon [Internet]. 2010 Dec [cited 2018 Sep 21];56(7):1103–12. Available from: http://linkinghub.elsevier.com/re trieve/pii/S0041010110000279 17.Berger M, Beys-da-Silva WO, Santi L, de Oliveira IM, Jorge PM, Henriques JAÔP, et al. Acute Lonomia obliqua caterpillar envenomationinduced physiopathological alterations in rats: Evidence of new toxic venom activities and the efficacy of serum therapy to counteract systemic tissue damage. Toxicon [Internet]. 2013;74:179–92. Available from: http://dx.doi.org/10.1016/j.toxic on.2013.08.061

18. Zager RA, Johnson ACM, Becker K. Renal cortical hemopexin accumulation in response to acute kidney injury. Am J Physiol Physiol [Internet]. 2012 Nov 15 [cited 2018 Nov 25];303(10):F1460–72. Available from: http://www.physiology.org/doi/1 0.1152/ajprenal.00426.2012

19. Berger M, Ribeiro Vieira MA, Almeida J. Acute kidney injury induced by snake and arthropod venoms. In: Renal failure: the facts. InTech; 2012.

20. Seibert CS, Tanaka-Azevedo AM, Santoro ML, Mackessy SP, Soares Torquato RJ, Lebrun I, et al. Purification of a phospholipase A2 from *Lonomia obliqua* caterpillar bristle extract. Biochem Biophys Res Commun [Internet]. 2006 Apr 21 [cited 2018 Nov 25];342(4):1027–33. Available from: http://www.ncbi.nlm.n ih.gov/pubmed/16510123

21. Linardi A, Rocha e Silva TAA, Miyabara EH, Franco-Penteado CF, Cardoso KC, Boer PA, et al. Histological and functional renal alterations caused by *Bothrops alternatus* snake venom: Expression and activity of Na+/K+-ATPase. Biochim Biophys Acta - Gen Subj [Internet]. 2011 Sep [cited 2018 Nov 25];1810(9):895–906. Available from: http://linkinghub.elsevier.com/re trieve/pii/S0304416511001334

22. Bohrer CB, Reck Junior J, Fernandes D, Sordi R, Guimarães JA, Assreuy J, et al. Kallikreinkinin system activation by *Lonomia obliqua* caterpillar bristles: involvement in edema and hypotension responses to envenomation. Toxicon. 2007 Apr;49(5):663–9.

23. Zannin M, Lourenço DM, Motta G, Dalla Costa LR, Grando M, Gamborgi GP, et al. Blood coagulation and fibrinolytic factors in 105 patients with hemorrhagic syndrome caused by accidental contact with *Lonomia obliqua* caterpillar in Santa Catarina, southern Brazil. Thromb Haemost [Internet]. 2003 Feb [cited 2018 Sep 21];89(2):355–64. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/12574817

24. Nascimento-Silva V, Rodrigues da Silva G, Moraes JA, Cyrino FZ, Seabra SH, Bouskela E, et al. A pro-inflammatory profile of endothelial cell in *Lonomia obliqua* envenomation. Toxicon [Internet]. 2012 Jul [cited 2018 Mar 19];60(1):50–60. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/22779081

25. Barrios M, Taylor P, Rodríguez-Acosta A, Sánchez EE, Arocha-Piñango CL, Gil A, et al. A mouse model to study the alterations in haemostatic and inflammatory parameters induced by Lonomia achelous caterpillar haemolymph. Toxicon [Internet]. 2012 Apr [cited 2018 Mar 19];59(5):547–54. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/22310207

26. Moraes J, Rodrigues G, Nascimento-Silva V, Renovato-Martins M, Berger M, Guimarães J, et al. Effects of *Lonomia obliqua* venom on vascular smooth muscle cells: contribution of NADPH oxidasederived reactive oxygen species. Toxins (Basel) [Internet]. 2017 Nov 7 [cited 2018 Nov 25];9(11):360. Available from: http://www.mdpi.com/2072-665 1/9/11/360

27. Pawluczyk IZ, Harris KP. Cytokine interactions promote synergistic fibronectin accumulation by mesangial cells. Kidney Int [Internet]. 1998 Jul 1 [cited 2018 Nov 25];54(1):62–70. Available from: http://www.ncbi.nlm.n ih.gov/pubmed/9648064

28. Veiga ABG, Pinto AFM, Guimarães JA. Fibrinogenolytic and procoagulant activities in the hemorrhagic syndrome caused by *Lonomia obliqua* caterpillars. Thromb Res [Internet]. 2003 [cited 2018 Sep 30];111(1–2):95–101. Available from: http://www.ncbi.nlm.n ih.gov/pubmed/14644086

29. Carrijo-Carvalho LC, Chudzinski-Tavassi AM. The venom of the Lonomia caterpillar: An overview. Toxicon [Internet]. 2007 May [cited 2018 Sep 30];49(6):741–57. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/17320134

30. Maggi S, Faulhaber GAM, Maggi S, Faulhaber GAM. *Lonomia obliqua* Walker (Lepidoptera: Saturniidae): hemostasis implications. Rev Assoc Med Bras [Internet]. 2015 Jun [cited 2018 Mar 19];61(3):263–8. Available from: http://www.scielo.br/scielo.php?s cript=sci\_arttext&pid=S0104-42302 015000300263&lng=en&tlng=en

31. Coll-Sangrona E, Arocha-Piñango CL. Fibrinolytic action on fresh human clots of whole body extracts and two semipurified fractions from Lonomia achelous caterpillar. Brazilian J Med Biol Res. 1998;31(6):779–84.

32. Alvarez Flores MP, Fritzen M, Chudzinski-Tavassi AM. Effects of LOSAC (a Factor X activator from Lonomia oblique caterpillar) on HUVECs and sequencing. Paper proceedings of Congresso da Sociedade Brasileira de Toxinologia, Andra dos Reis; 2004. 33. Fritzen M, Alvarez Flores MP, Reis CV, Chudzinski-Tavassi AM. A prothrombin activator (Lopap) modulating inflammation, coagulation cell survival mechanisms. and Biochem Biophys Res Commun [Internet]. 2005 Jul 29 [cited 2018 Mar 19];333(2):517–23. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/15963468

34. Fritzen M, Schattner M, Quintana Ribeiro AL, de Fátima Correia Batista I, Ventura J, Prezoto BC, et al. *Lonomia obliqua* venom action on fibrinolytic system. Thromb Res [Internet]. 2003 Jan [cited 2018 Sep 30];112(1–2):105–10. Available from: http://linkinghub.elsevier.com/re trieve/pii/S0049384803005863

35. Bastos L de С, Veiga ABG, Guimarães JA, Tonussi CR. Nociceptive and edematogenic responses elicited by a crude bristle extract of Lonomia obliqua caterpillars. Toxicon [Internet]. 2004 Mar 1 [cited 2018 Sep 30];43(3):273-8. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/15033325

36. Burdmann EA, Antunes I, Saldanha LB, Abdulkader RC. Severe acute renal failure induced by the venom of Lonomia caterpillars. Clin Nephrol [Internet]. 1996 Nov [cited 2018 Sep 30];46(5):337–9. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/8953124

37. Cárdenas P, Arbelbide J, Nucifora E, Otaso J, Barrera L. Coagulopatia grave por Lonomia. Hematologia [Internet].
2002 [cited 2018 Sep 30];6(2).
Available from: http://www.sah.org.ar/ revista/numeros/vol6-n2-36-41-.pdf

38. Rocha-Campos AC, Gonçalves LR, Higashi HG, Yamagushi IK, Fernandes I, Oliveira JE, et al. Specific heterologous F(ab')2 antibodies revert blood incoagulability resulting from envenoming by *Lonomia obliqua* caterpillars. Am J Trop Med Hyg [Internet]. [cited 2018 Sep 30];64(5– 6):283–9. Available from: http://www. ncbi.nlm.nih.gov/pubmed/11463118

39. Arocha-Piñango CL, Marval E, Guerrero B. Lonomia genus caterpillar toxins: biochemical aspects. Biochimie [Internet]. 2000 [cited 2018 Sep 30];82(9–10):937–42. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/11086223

40. Fundação Nacional de Saúde, Ministério da Saúde. Manual de diagnóstico e tratamento de acidentes por animais peçonhentos [Internet]. 2nd ed. Brasilia; 2001 [cited 2018 Sep 30]. Available from: http://bvsms.saude.gov.br/bvs/pu blicacoes/funasa/manu\_peconhentos.p df

41. Bogdanov VY, Balasubramanian V, Hathcock J, Vele O, Lieb M, Nemerson Y. Alternatively spliced human tissue factor: a circulating, soluble, thrombogenic protein. Nat Med [Internet]. 2003 Apr 24 [cited 2018 Sep 30];9(4):458–62. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/12652293

42. Peña-Vásquez W, Vásquez-Paz H, Vásquez-Becerra R, Chiappe-Gonzalez A, Ñavincopa-Flores M, Ticona-Chávez E. Niña con erucismo hemorrágico por *Lonomia* spp. reporte de un caso. Rev Peru Med Exp Salud Publica [Internet]. 2016 Dec 13 [cited 2018 Sep 30];33(4):819. Available from: http://www.ncbi.nlm.nih.gov/pu bmed/28327855

43. Lucena S, Salazar AM, Gil A, Arocha-Piñango CL, Guerrero B. The action of Lonomin V (Lonomia achelous) on fibronectin functional properties. Thromb Res [Internet]. 2008 [cited 2018 Sep 30];121(5):653– DANIEL MARTÍN ARSANIOS, ELÍAS JOSUÉ QUINTERO MUÑOZ, NICOLÁS DAVID SANTOYO SARMIENTO, ET AL.

61. Available from: http://www.ncbi.nl m.nih.gov/pubmed/17997475
44. Arocha-Piñango CL, Guerrero B. Síndrome hemorrágico producido por contacto con orugas. Estudios clínicos y experimentales. Revisión. Invest Clin. 2003;44(2):155–63.

## Notes

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